

Synthesis of 6-amino-1,6-anhydro-6-deoxysugar derivatives

Dominique Lafont *, Andreas Wollny¹, Paul Boullanger

Laboratoire de Chimie Organique II, Unité Mixte de Recherche CNRS 5622, Université de Lyon 1, Chimie Physique Electronique de Lyon, 43 Bd du 11 Novembre 1918, F-69622 Villeurbanne, France

Received 2 February 1998; accepted 27 March 1998

Abstract

Staudinger reaction of triphenylphosphine with 2,3,4-tri-*O*-acetyl-6-*O*-*p*-tolylsulfonyl- β -D-glycopyranosyl azides led to an anomeric iminophosphorane which rearranged in situ by elimination of the sulfonate at C-6. The 1,6-anhydro-6-deoxy-6-triphenylphosphonioamino- β -D-glycopyranose salts thus obtained were transformed into the corresponding 2,3,4-tri-*O*-acetyl-6-amino-1,6-anhydro-6-deoxy- β -D-glycopyranoses which were further *N*-acylated or *N*-alkoxycarbonylated. ¹H and ¹³C NMR of these products show the occurrence of two rotamers in solution, due to restricted rotations around the amide bond. © 1998 Elsevier Science Ltd. All rights reserved

Keywords: 6-*O*-*p*-tolylsulfonyl- β -D-glycosyl azide ; Staudinger reaction ; 6-Amino-1,6-anhydro-6-deoxy- β -D-glycopyranose derivatives

1. Introduction

Only a few examples of 6-amino-1,6-anhydro-6-deoxy sugars have been described in the literature. Thus, treatment of 6-amino-6-deoxy-1,2-*O*-isopropylidene- β -L-idofuranose in acidic conditions led to an acyclic derivative, which was transformed under basic conditions into 6-amino-6-deoxy-L-idopyranose. The latter underwent a spontaneous and practically quantitative dehydration into 6-amino-1,6-anhydro-6-deoxy- β -L-idopyranose [1–4]. The anhydro ring closure was evidently favored by the stereochemistry of the hydroxyl groups in equatorial position. Under the same conditions, a similar transformation was not observed in the

D-*gluco* and D-*galacto* series since, in this case, a conformational inversion would be necessary to obtain the anhydro derivative [4]. Recently, Pradera et al. showed that a 6-amino-1,6-anhydro-6-deoxy-2,3,4-tri-*O*-mesyl- β -D-glucopyranose derivative could be prepared by treatment of *N*-diethoxycarbonylvinyl-2,3,4,6-tetra-*O*-mesyl- β -D-glucopyranosylamine under basic conditions [5]. The attack of the nitrogen atom was regiospecific at the 6-position. With 2,3,6-tri-*O*-acyl-*N*-diethoxycarbonylvinyl-4-*O*-mesyl-glycopyranosylamines, syntheses of *O*-protected 4-deoxy-4-diethoxycarbonylvinylaminoaldoses were also possible, through nucleophilic substitution of the mesyloxy group at C-4 with inversion of configuration [6].

In our previous papers on β -glycosylation in the 2-amino-2-deoxy-D-glucose and lactosamine series, starting from 1,2-*trans*-2-deoxy-2-iodo- α -D-glyco-

* Corresponding author.

¹ On leave from the University of Stuttgart.

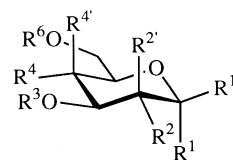
pyranosyl azides [7,8], β -glycosides were obtained by the Staudinger reaction of the azido function with triphenylphosphine [9]. Nucleophilic attack of the nitrogen at C-2 led to an unstable aziridine intermediate, which was opened by the alcohol at the anomeric position. A further application of the Staudinger reaction towards the synthesis of 6-amino-1,6-anhydro-6-deoxysugar derivatives (*D*-gluco, *D*-galacto and *D*-manno series) is reported in the present paper.

2. Results and discussion

Reaction of azido sugars or glycosyl azides and their corresponding imino-phosphoranes has already been reported in the literature [8,10–15]. Due to their ylide structure, in which the nitrogen atom is negatively charged, sugar iminophosphoranes are strong nucleophiles which could intramolecularly substitute a 6-*O*-*p*-tolylsulfonyl group in the same way as an anomeric hydroxyl group [16] or mercapto group [17], leading to 1,6-anhydro derivatives.

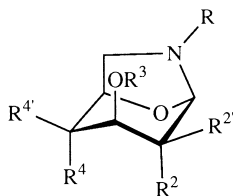
Therefore, 2,3,4-tri-*O*-acetyl-6-*O*-*p*-tolylsulfonyl- β -D-glucopyranosyl azides **3** and **4** were prepared in two steps from 1,2,3,4-tetra-*O*-acetyl-6-*O*-*p*-tolylsulfonyl-D-glucopyranose **1** [18] and **2** [19], by treatment with 33% hydrogen bromide in acetic acid and transformation of the glycosyl bromides into glycosyl azides by phase transfer catalysis in the presence of sodium azide according to Roy et al. [20]. 2,3,4-Tri-*O*-acetyl-6-*O*-*p*-tolylsulfonyl- β -D-mannopyranosyl azide **6** was synthesized in three steps from 2,3,4,6-tetra-*O*-benzoyl- β -D-mannopyranosyl azide **5** [21], (i.e. debenzoylation according to Zemplén, regioselective tosylation of the 6-OH and acetylation) in a 56% overall yield. Treatment of the azides **3**, **4** or **6** with a slight excess of triphenylphosphine (1.05 equiv) in dichloromethane afforded quantitatively the expected 2,3,4-tri-*O*-acetyl-1,6-anhydro-6-deoxy-6-triphenylphosphonioamino- β -D-glycopyranose *p*-tolylsulfonates **7**, **8** and **9**. The structure of these salts was evidenced from ^1H -, ^{13}C - and ^{31}P NMR spectra; thus, ^1H NMR couplings values were characteristic of a $^1\text{C}_4$ (*D*) conformation as in 1,6-anhydrosugar derivatives [22,23], and the presence of the C-6–N–C-1 bridge was ascertained by ^1H NMR couplings $J_{\text{H,P}}$ ($J_{1,\text{P}}$ 8.3–8.7 Hz, $J_{6b,\text{P}}$ 10–13 Hz) and ^{13}C chemical shift for C-6 (δ 48.34–49.67 ppm). The formation of

phosphonioamino salts was also supported by the ^{31}P chemical shifts (δ 43.26–45.69) and by the coupling constants $J_{\text{C,P}}$ for C_{ipso} , C_{ortho} , C_{meta} and C_{para} of the triphenyl-phosphonio moiety [8,24,25]. The salts **7–9** were transformed into the crude 3,4,6-tri-*O*-acetyl-6-amino-1,6-anhydro-6-deoxy- β -D-glycopyranoses **10–12** by anionic exchange of the anion [Dowex 2X8 (OH^-) column in dichloromethane] via the phosphonioamino hydroxide derivative, which rearranged immediately, affording triphenylphosphine oxide as by-product. These derivatives were not isolable by column chromatography and were used directly for *N*-protection reactions. Furthermore, ^1H NMR of **12** displayed a second *N*-acetylated product **13**, resulting from the $\text{O} \rightarrow \text{N}$ migration of the O-2 acetyl group. This was confirmed by peracetylation of the mixture, which afforded **16** only. ^1H and ^{13}C NMR signals of derivatives **10** and **11** were fully assigned at 500 MHz in hexadeuteroacetone, using NOESY and ^1H – ^{13}C 2D experiments. ROESY [26] was also necessary for **10** in which most of the vicinal and long-range coupling constants were of the same order of magnitude [22,23].



- 1 $\text{R}^1, \text{R}^1' = \text{OAc}, \text{H}, \text{R}^2 = \text{R}^4 = \text{OAc}, \text{R}^3 = \text{Ac}, \text{R}^6 = \text{Ts}, \text{R}^2' = \text{R}^4' = \text{H}$
- 2 $\text{R}^1, \text{R}^1' = \text{OAc}, \text{H}, \text{R}^2 = \text{R}^4' = \text{OAc}, \text{R}^3 = \text{Ac}, \text{R}^6 = \text{Ts}, \text{R}^2' = \text{R}^4 = \text{H}$
- 3 $\text{R}^1' = \text{N}_3, \text{R}^2 = \text{R}^4' = \text{OAc}, \text{R}^3 = \text{Ac}, \text{R}^6 = \text{Ts}, \text{R}^1 = \text{R}^2' = \text{R}^4' = \text{H}$
- 4 $\text{R}^1' = \text{N}_3, \text{R}^2 = \text{R}^4' = \text{OAc}, \text{R}^3 = \text{Ac}, \text{R}^6 = \text{Ts}, \text{R}^1 = \text{R}^2' = \text{R}^4 = \text{H}$
- 5 $\text{R}^1' = \text{N}_3, \text{R}^2' = \text{R}^4' = \text{OBz}, \text{R}^3 = \text{R}^6 = \text{Bz}, \text{R}^1 = \text{R}^2 = \text{R}^4' = \text{H}$
- 6 $\text{R}^1' = \text{N}_3, \text{R}^2' = \text{R}^4' = \text{OAc}, \text{R}^3 = \text{Ac}, \text{R}^6 = \text{Ts}, \text{R}^1 = \text{R}^2 = \text{R}^4' = \text{H}$

N-Acylation of the crude derivatives **10,11** and the mixture **12,13** (pyridine/acetic anhydride) gave the pure 6-acetamido-2,3,4-tri-*O*-acetyl-1,6-anhydro-6-deoxy- β -D-glycopyranoses **14–16** in high yields. ^1H NMR spectra of **14,15** and **16** in deuteriochloroform revealed the presence of two conformers, the proportions of which were dependent of their structures (**14**: 52/48, **15**: 55/45, **16**: 43/57). This could be due to restricted rotations around the amide bond [1,27–30]. On increasing the temperature, in hexadeuterodimethyl-sulfoxide, the signals coalescence was observed above 115 °C for compound **14**. The presence of two rotamers was also confirmed by the ^{13}C NMR spectra of derivatives **14–16**.



- 7 $R^2 = R^4 = \text{OAc}$, $R^{2'} = R^{4'} = \text{H}$, $R^3 = \text{Ac}$, $R = \text{PPh}_3^+$, TsO^-
- 8 $R^2 = R^4 = \text{OAc}$, $R^{2'} = R^{4'} = \text{H}$, $R^3 = \text{Ac}$, $R = \text{PPh}_3^+$, TsO^-
- 9 $R^{2'} = R^4 = \text{OAc}$, $R^2 = R^{4'} = \text{H}$, $R^3 = \text{Ac}$, $R = \text{PPh}_3^+$, TsO^-
- 10 $R^2 = R^4 = \text{OAc}$, $R^{2'} = R^{4'} = \text{R}$, $R^3 = \text{Ac}$
- 11 $R^2 = R^4 = \text{OAc}$, $R^{2'} = R^{4'} = \text{R}$, $R^3 = \text{Ac}$
- 12 $R^{2'} = R^4 = \text{OAc}$, $R^2 = R^{4'} = \text{R}$, $R^3 = \text{Ac}$
- 13 $R^{2'} = \text{OH}$, $R^4 = \text{OAc}$, $R^2 = R^{4'} = \text{H}$, $R = \text{Ac}$
- 14 $R^2 = R^4 = \text{OAc}$, $R^{2'} = R^{4'} = \text{H}$, $R = R^3 = \text{Ac}$
- 15 $R^2 = R^4 = \text{OAc}$, $R^{2'} = R^{4'} = \text{H}$, $R = R^3 = \text{Ac}$
- 16 $R^{2'} = R^4 = \text{OAc}$, $R^2 = R^{4'} = \text{H}$, $R = R^3 = \text{Ac}$
- 17 $R^2 = R^4 = \text{OAc}$, $R^{2'} = R^{4'} = \text{H}$, $R^3 = \text{Ac}$, $R = \text{COOMe}$
- 18 $R^2 = R^4 = \text{OAc}$, $R^{2'} = R^{4'} = \text{H}$, $R^3 = \text{Ac}$, $R = \text{COOAl}$
- 19 $R^2 = R^4 = \text{OAc}$, $R^{2'} = R^{4'} = \text{H}$, $R^3 = \text{Ac}$, $R = \text{COC}_{13}\text{H}_{27}$
- 20 $R^2 = R^4 = \text{OAc}$, $R^{2'} = R^{4'} = \text{H}$, $R^3 = \text{Ac}$, $R = \text{COC}_7\text{H}_{15}$
- 21 $R^2 = R^4 = \text{OH}$, $R^{2'} = R^{4'} = \text{R}^3 = \text{H}$, $R = \text{COC}_{13}\text{H}_{27}$
- 22 $R^2 = R^4 = \text{OH}$, $R^{2'} = R^{4'} = \text{H}$, $R^3 = \text{H}$, $R = \text{COC}_7\text{H}_{15}$

Since 6-*N*-acyl-6-amino-1,6-anhydro-6-deoxy- β -D-glycopyranoses could find applications in the field of detergents, liquid crystals or surface-active complexing agents, crude compound **10** was alkoxycarbonylated or acylated. Thus, reaction of **10** in pyridine with an excess of alkyl chloroformate (methyl or allyl chloroformate) or acyl chloride (octanoyl or tetradecanoyl chloride) afforded the expected carbamates **17,18** and amides **19** and **20** in good to high yields. As for acetamide derivatives, ^1H and ^{13}C NMR spectra of the carbamates also displayed the presence of two rotamers at room temperature. Product **19** and **20** were deacetylated by the Zemplén procedure, and solubility in water of the deprotected sugars **21** and **22** was determined. 1,6-Anhydro-6-deoxy-6-tetradecanoyl-amino- β -D-glucopyranose (**21**) was insoluble in water, contrary to 1,6-anhydro-6-deoxy-6-octanoylamino- β -D-glucopyranose (**22**). Krafft temperature of **22** was shown to be below 20 °C and the surface tension (γ) measurements were carried out by the ring method of Lecomte du Nouÿ [31] and corrected according to Harkins and Jordan [32]. Measurements were performed at room temperature (20 °C) and the cmc was determined at the break of the slope in the γ versus $\log[\text{C}]$ plots, as usual. The interfacial area per molecule (ao) was calculated for concentrations just below the cmc according to the Gibbs law. The experimental values thus determined were, respec-

tively, 0.019 mM for the cmc, 30.5 mN·m⁻¹ for γ at the interface and 39 Å² for ao . The cmc value was very close to that found for octyl β -D-glucopyranoside [33], but ao was smaller (49 Å² for octyl β -D-glucopyranoside [34]), which could indicate a lower hydration of the carbohydrate head.

In conclusion, application of the Staudinger reaction to 6-*O*-*p*-tolylsulfonyl-2,3,4-tri-*O*-acetyl- β -D-glycopyranosyl azides allowed the formation of 6-amino-1,6-anhydro-6-deoxy- β -D-glycopyranose derivatives, which could be *N*-alkoxycarbonylated or *N*-acylated. This methodology is short and efficient, and well suited for the preparation of new surfactants.

2. Experimental

General methods.—Pyridine was dried by boiling with CaH₂ prior to distillation. Dichloromethane was washed twice with water, dried with CaCl₂ and distilled from CaH₂. Methanol was refluxed with NaOMe before distillation. Pyridine and CH₂Cl₂ were stored over 4 Å molecular sieves and MeOH over 3 Å molecular sieves. Melting points were determined on a Büchi apparatus and were uncorrected. Thin layer chromatography was performed on aluminium sheets coated with Silica Gel 60 F₂₅₄ (E. Merck). Compounds were visualized by spraying the TLC plates with dilute 15% aq H₂SO₄, followed by charring at 150 °C for a few min. Column chromatography was performed on Silica-gel Geduran Si 60 (E. Merck). Optical rotations were recorded on a Perkin–Elmer 241 polarimeter in a 1 dm cell at 21 °C. ^1H and ^{13}C NMR spectra were recorded with Bruker AC-200, AM-300 or DRX-500 spectrometers working at 200, 300 or 500 MHz and 50, 75.5 or 125 MHz, respectively, with Me₄Si as internal standard. Elemental analyses were performed by the “Laboratoire Central d’Analyses du CNRS” (Vernaison, France).

2,3,4-Tri-*O*-acetyl-6-*O*-*p*-tolylsulfonyl- β -D-glucopyranosyl azide (3**).**—1,2,3,4-Tetra-*O*-acetyl-6-*O*-*p*-tolylsulfonyl- α,β -D-glucopyranose (**1**) [18] (4.50 g, 8.95 mmol) in CH₂Cl₂ (5 mL) was treated with 33% HBr in AcOH (12 mL). After 1.5 h, the mixture was concentrated, the residue was dissolved in CHCl₃ (100 mL), the organic phase was washed with cooled aq NaHCO₃, then with water, dried and concentrated again. To a soln of this glucosyl bromide (8.95 mmol), Bu₄NHSO₄ (3.04 g, 8.95 mmol, 1 equiv.) and NaN₃ (2.33 g, 4 equiv.) in

CH_2Cl_2 (50 mL), was added satd aq NaHCO_3 (50 mL); the two phase mixture was vigorously stirred for 2 h; addition of EtOAc (450 mL), washing of the organic phase with aq NaHCO_3 (2×100 mL), then with brine (100 mL), drying and concentration afforded the crude product **2** which was recrystallized twice from absolute EtOH : 3.13 g, yield 72%; mp 129–130 °C (lit [35]. mp 128–130 °C); $[\alpha]_{\text{D}} + 19.0^\circ$ (c 1.0, CHCl_3); R_f 0.70 (1:1 EtOAc –petroleum ether).

2,3,4-Tri-O-acetyl-6-O-tosyl- β -D-galactopyranosyl azide (4).—Prepared in 75% yield, as described previously from 1,2,3,4-tetra-O-acetyl-6-O-tosyl- α,β -D-galactopyranose (**2**) [19]; mp 94 °C (EtOH) [lit [35] mp 92–94 °C (diethyl ether)]; $[\alpha]_{\text{D}} - 16.3^\circ$ (c 1.0, CHCl_3); R_f 0.70 (1:1 EtOAc –petroleum ether).

2,3,4-Tri-O-acetyl-6-O-p-tolylsulfonyl- β -D-mannopyranosyl azide (6).—2,3,4,6-Tetra-O-benzoyl- β -D-mannopyranosyl azide (**5**) [21] (4.10 g, 6.60 mmol) was treated overnight in MeOH (50 mL) containing a catalytic amount of sodium; after concentration, the residue was washed with petroleum ether (3×30 mL), dissolved in $\text{C}_5\text{H}_5\text{N}$ (12 mL) and cooled to -12°C ; a soln of TsCl (1.67 g, 8.76 mmol, 1.32 equiv) in $\text{C}_5\text{H}_5\text{N}$ (6 mL) was added dropwise for 1 h and the mixture was allowed to reach room temperature. Stirring was maintained overnight, then the soln was concentrated and the residue was purified by column chromatography (EtOAc) affording the 6-O-monosulfonate as a syrup (1.66 g, 70% yield). The latter was immediately acetylated in a 2:1 mixture of $\text{C}_5\text{H}_5\text{N}$ and Ac_2O (15 mL). After 16 h, concentration of the soln and column chromatography of the residue (1:1 EtOAc –petroleum ether) afforded the pure product **6** (1.79 g, 56% overall yield) as a crystalline solid: mp 128–129 °C; $[\alpha]_{\text{D}} + 43.0^\circ$ (c 1.0, CHCl_3); R_f 0.67 (1:1 EtOAc –petroleum ether); ^1H NMR (CDCl_3): δ 7.80 and 7.34 (2d, 4 H, J 8.2 Hz, CH_3 -Ph), 5.41 (dd, 1 H, $J_{1,2}$ 1.2 Hz, $J_{2,3}$ 3.1 Hz, H-2), 5.15 (dd, 1 H, $J_{3,4}$ 10.1 Hz, $J_{4,5}$ 9.7 Hz, H-4), 5.01 (dd, 1 H, H-3), 4.64 (d, 1 H, H-1), 4.15 (d, 2 H, $J_{5,6a}$ 4.5 Hz, $J_{5,6b}$ 4.5 Hz, H-6a,6b), 3.80 (dt, 1 H, H-5), 2.45 (s, 3 H, CH_3Ph), 2.17, 2.05, 1.98 (3s, 9 H, CH_3COO); ^{13}C NMR (CDCl_3): δ 169.87, 169.83, 169.63 (3 C, CH_3COO), 145.22 (CSO_2), 132.42 (CCH_3), 129.96 and 128.06 (4 C, PhSO_2), 84.85 (C-1), 74.46, 70.72, 69.21, 65.44 (C-2,3,4,5), 67.91 (C-6), 21.66 (CH_3Ph), 20.67, 20.59, 20.48 (3 C, CH_3COO). Anal. Calcd for $\text{C}_{19}\text{H}_{23}\text{N}_3\text{O}_{10}\text{S}$ (485.46): C, 47.00; H, 4.78; N, 8.66. Found: C, 46.97; H, 4.91, N, 8.78.

General procedure for the preparation of the cyclic 1,6-anhydro-6-deoxy-6-triphenylphosphonioamino p-tolylsulfonates (7–9).—A soln of PPh_3 (0.57–1.14 g, 2.17–4.34 mmol) in CH_2Cl_2 (2–4 mL) was added dropwise under an argon atmosphere, to a soln of the peracetylated 6-O-p-tolylsulfonyl glycosylazides **3**, **4** or **6** (1.00–2.00 g, 2.06–4.12 mmol) in dry CH_2Cl_2 (6–12 mL). Evolution of nitrogen started after a few minutes and the soln was stirred overnight. After concentration, the ^1H NMR spectrum of the residue showed the presence of the expected product besides some traces of PPh_3 and PPh_3O , which could be removed by three successive washings with CH_2Cl_2 (1 mL) and petroleum ether (10 mL).

2,3,4-Tri-O-acetyl-1,6-anhydro-6-deoxy-6-triphenylphosphonioamino- β -D-glucopyranose p-tolylsulfonate (7).—Obtained quantitatively as described above from 2,3,4-tri-O-acetyl-6-O-p-tolylsulfonyl- β -D-glucopyranosyl azide (**3**) (2.00 g, 4.12 mmol). Amorphous hygroscopic solid; $[\alpha]_{\text{D}} - 12.9^\circ$ (c 1.0, CHCl_3); ^1H NMR (CD_3COCD_3): δ 8.08–7.85 (m, 15 H, 3 Ph), 7.64 and 7.02 (2d, 4 H, J 8.2 Hz, CH_3 -Ph), 5.67 (ddd, 1 H, $J_{1,2}$, $J_{1,3}$ 1.4 Hz, $J_{1,P}$ 8.3 Hz, H-1), 5.04 (dddd, 1 H, $J_{3,5}$, $J_{4,5}$ 1.4 Hz, $J_{5,6a}$ 1.7 Hz, $J_{5,6b}$ 6.8 Hz, H-5), 4.88 (dddd, 1 H, $J_{3,4}$, $J_{2,3}$ 1.4 Hz, H-3), 4.83 (dd, 1 H, H-4), 4.50 (dd, 1 H, H-2), 3.92 (dd, 1 H, $J_{6a,6b}$ 9.7 Hz, H-6a), 3.84 (ddd, 1 H, $J_{6b,P}$ 10.5 Hz, H-6b), 2.27 (s, 3 H, CH_3Ph), 2.20, 2.10, 1.97 (3s, 9 H, CH_3COO); ^{31}P NMR (CDCl_3): δ 43.69; ^{13}C NMR (CDCl_3): δ 169.11, 169.09, 168.46 (3 C, CH_3COO), 144.55 (CSO_2), 137.92 (CCH_3), 136.06 (d, 3 C, $J_{C,P}$ 2.7 Hz, C_{para}), 133.47 (d, 6 C, $J_{C,P}$ 11.1 Hz, C_{ortho}), 130.70 (d, 6 C, $J_{C,P}$ 14.3 Hz, C_{meta}), 127.92 and 125.92 (4 C, PhSO_2), 117.88 (d, 3 C, $J_{C,P}$ 101.9 Hz, C_{ipso}), 86.45 (d, $J_{C1,P}$ 3.2 Hz, C-1), 76.19 (d, $J_{C5,P}$ 6.1 Hz, C-5), 68.71 (C-4), 68.57 (d, $J_{C2,P}$ 6.5 Hz, C-2), 68.25 (C-3), 49.05 (C-6), 21.01 (PhCH_3), 20.68, 20.61, 20.37 (3 C, CH_3COO). Anal. Calcd for $\text{C}_{37}\text{H}_{38}\text{NO}_{10}\text{PS}$, $2\text{H}_2\text{O}$ (755.76): C, 58.80; H, 5.60; N, 1.85. Found: C, 58.87; H, 5.44, N, 1.92.

2,3,4-Tri-O-acetyl-1,6-anhydro-6-deoxy-6-triphenylphosphonioamino- β -D-galactopyranose p-tolylsulfonate (8).—Obtained quantitatively as described above from 2,3,4-tri-O-acetyl-6-O-p-tolylsulfonyl- β -D-galactopyranosyl azide (**4**) (2.00 g, 4.12 mmol). Amorphous solid; $[\alpha]_{\text{D}} + 14.0^\circ$ (c 1.0, CHCl_3); ^1H NMR (CD_3COCD_3): δ 8.08–7.85 (m, 15 H, 3 Ph), 7.64 and 7.02 (2d, 4 H, J 8.2 Hz, CH_3 -Ph), 5.62 (ddd, 1 H, $J_{1,2}$, $J_{1,3}$ 1.4 Hz, $J_{1,P}$ 8.4 Hz, H-1), 5.34 (dddd, 1 H, $J_{2,3}$, $J_{3,5}$ 1.4 Hz, $J_{3,4}$ 5.1 Hz, H-3), 5.24

(dd, 1 H, $J_{4,5}$ 4.4 Hz H-4), 5.02 (m, 1 H, H-5), 4.72 (dd, 1 H, H-2), 3.87 (dd, 1 H, $J_{5,6a}$ 1.5 Hz, $J_{6a,6b}$ 7.8 Hz, H-6a), 3.82 (ddd, 1 H, $J_{5,6b}$ 5.8 Hz, $J_{6b,P}$ 9.0 Hz, H-6b), 2.27 (s, 3 H, CH_3Ph), 2.18, 1.99, 1.95 (3s, 9 H, CH_3COO); ^{31}P NMR (CDCl_3): δ 43.26; ^{13}C NMR (CDCl_3): δ 169.08, 169.01, 168.96 (3 C, CH_3COO), 144.72 (CSO_2), 137.95 (CCH_3), 136.13 (d, 3 C, $J_{C,P}$ 2.9 Hz, C_{para}), 133.58 (d, 6 C, $J_{C,P}$ 11.1 Hz, C_{ortho}), 130.38 (d, 6 C, $J_{C,P}$ 13.2 Hz, C_{meta}), 127.93 and 125.98 (4 C, PhSO_2), 117.92 (d, 3 C, $J_{C,P}$ 102.0 Hz, C_{ipso}), 86.40 (d, $J_{C1,P}$ 3.3 Hz, C-1), 74.55 (d, $J_{C5,P}$ 6.3 Hz, C-5), 71.00 (d, $J_{C2,P}$ 7.0 Hz, C-2), 67.03 (C-3), 63.94 (C-4), 48.34 (C-6), 21.05 (PhCH_3), 20.54, 20.35, 20.33 (3 C, CH_3COO). Anal. Calcd for $\text{C}_{37}\text{H}_{38}\text{NO}_{10}\text{PS}$, $2\text{H}_2\text{O}$ (755.76): C, 58.80; H, 5.60; N, 1.85. Found: C, 58.53; H, 5.38, N, 1.66.

2,3,4-Tri-O-acetyl-1,6-anhydro-6-deoxy-6-triphenylphosphonioamino- β -D-mannopyranose p-tolylsulfonate (9).—Obtained quantitatively as described above from 2,3,4-tri-O-acetyl-6-O-p-tolylsulfonfyl- β -D-mannopyranosyl azide (**6**) (1.00 g, 2.06 mmol). Amorphous solid; $[\alpha]_D -28.2^\circ$ (c 1.0, CHCl_3); ^1H NMR (CD_3COCD_3): δ 8.12–7.85 (m, 15 H, 3 *Ph*), 7.63 and 7.02 (2d, 4 H, J 8.0 Hz, $\text{CH}_3\text{-Ph}$), 5.78 (ddd, 1 H, $J_{1,2}$ 2.7 Hz, $J_{1,3}$ 0.7 Hz, $J_{1,P}$ 8.7 Hz, H-1), 5.35 (dddd, 1 H, $J_{2,3}$ 5.2 Hz, $J_{3,4}$, $J_{3,5}$ 1.4 Hz, H-3), 5.09 (dddd, 1 H, $J_{4,5}$ 1.4 Hz, $J_{5,6a}$ 2.7 Hz, $J_{5,6b}$ 7.2 Hz, H-5), 5.03 (dd, 1 H, H-2), 5.01 (m, 1 H, H-4), 4.05 (ddd, 1 H, $J_{4,6a}$ 1.4 Hz, $J_{6a,6b}$ 9.9 Hz, H-6a), 3.84 (ddd, 1 H, $J_{6b,P}$ 13.2 Hz, H-6b), 2.37 (s, 3 H, CH_3Ph), 2.27, 2.13, 1.70 (3s, 9 H, CH_3COO); ^{31}P NMR (CDCl_3): δ 45.69; ^{13}C NMR (CDCl_3): δ 169.19, 169.10, 168.88 (3 C, CH_3COO), 144.70 (CSO_2), 137.83 (CCH_3), 135.94 (d, 3 C, $J_{C,P}$ 2.7 Hz, C_{para}), 133.63 (d, 6 C, $J_{C,P}$ 8.9 Hz, C_{ortho}), 130.51 (d, 6 C, $J_{C,P}$ 13.2 Hz, C_{meta}), 127.86 and 125.86 (4 C, PhSO_2), 118.22 (d, 3 C, $J_{C,P}$ 101.9 Hz, C_{ipso}), 86.64 (C-1), 75.66 (d, $J_{C5,P}$ 5.4 Hz, C-5), 71.09 (C-4), 66.45 (C-3), 66.14 (d, $J_{C2,P}$ 6.4 Hz, C-2), 49.67 (C-6), 21.92 (PhCH_3), 20.68, 20.59, 20.19 (3 C, CH_3COO). Anal. Calcd for $\text{C}_{37}\text{H}_{38}\text{NO}_{10}\text{PS}$, $2\text{H}_2\text{O}$ (755.76): C, 58.80; H, 5.60; N, 1.85. Found: C, 58.89; H, 5.36, N, 1.79.

General procedure for the preparation of the 6-acetamido-2,3,4-tri-O-acetyl-1,6-anhydro-6-deoxy- β -D-hexopyranoses (14–16).—A soln of the crude salts **7**, **8** or **9**, prepared from **3**, **4** or **6** (0.337 g, 0.695 mmol) and PPh_3 (0.191 g, 1.05 equiv in CH_2Cl_2 (1 mL), was applied at the top of a column of Dowex 2X8 (OH^-) in CH_2Cl_2 and eluted with the same solvent. Concentration of the eluate

afforded a mixture of the expected product **10** or **11** (from **7** or **8**) or **12/13** (from **9**) with PPh_3O . These compounds were too instable to be purified by column chromatography on silica-gel.

NMR spectroscopy of crude 2,3,4-tri-O-acetyl-6-amino-1,6-anhydro-6-deoxy- β -D-glucopyranose (10). ^1H ($\text{CD}_3\text{-COCD}_3$): δ 4.98 (ddd, 1 H, $J_{1,2}$, $J_{1,3}$ 1.4 Hz, $J_{1,5}$ 0.6 Hz, H-1), 4.78 (dddd, 1 H, $J_{2,3}$, $J_{3,4}$, $J_{3,5}$ 1.4 Hz, H-3), 4.54 (dddd, 1 H, $J_{2,4}$, $J_{4,5}$ 1.4 Hz, $J_{4,6b}$ 0.6 Hz, H-4), 4.50 (dddd, 1 H, $J_{5,6a}$ 7.0 Hz, $J_{5,6b}$ 1.5 Hz, H-5), 3.15 (dd, 1 H, $J_{6a,6b}$ 10.2 Hz, H-6a), 3.09 (ddd, 1 H, H-6b), 2.08, 2.06, 2.06 (3s, 9 H, CH_3COO); ^{13}C (CD_3COCD_3): δ 169.19, 169.10, 168.88 (3 C, CH_3COO), 88.05 (C-1), 74.03 (C-5), 72.95 (C-4), 72.90 (C-2), 71.07 (C-3), 46.07 (C-6), 20.68, 20.59, 20.19 (3 C, CH_3COO).

NMR spectroscopy of crude 2,3,4-tri-O-acetyl-6-amino-1,6-anhydro-6-deoxy- β -D-galactopyranose (11). ^1H ($\text{CD}_3\text{-COCD}_3$): δ 5.16 (ddd, 1 H, $J_{1,3}$, $J_{2,3}$ 1.4 Hz, $J_{3,4}$ 5.5 Hz, H-3), 5.08 (ddd, 1 H, $J_{4,5}$ 4.3 Hz, $J_{4,6b}$ 0.9 Hz, H-4), 4.98 (dd, 1 H, $J_{1,2}$ 1.4 Hz, H-1), 4.62 (dd, 1 H, H-2), 4.37 (ddd, 1 H, $J_{5,6a}$ 1.0 Hz, $J_{5,6b}$ 6.4 Hz, H-5), 3.39 (dd, 1 H, $J_{6a,6b}$ 9.8 Hz, H-6a), 3.03 (ddd, 1 H, H-6b), 2.15, 2.13, 2.03 (3s, 9 H, CH_3COO); ^{13}C (CD_3COCD_3): δ 170.23, 169.94, 169.79 (3 C, CH_3COO), 88.03 (C-1), 73.86 (C-2), 72.77 (C-5), 68.52 (C-3), 66.96 (C-4), 44.78 (C-6), 20.87, 20.78, 20.60 (3 C, CH_3COO).

Acetylation of the crude compounds **10**, **11** or **12/13** with a 2:1 pyridine– Ac_2O mixture (15 h) followed by concentration afforded products **14**, **15** or **16** which were purified by column chromatography (7:1 EtOAc–EtOH).

6-Acetamido-2,3,4-tri-O-acetyl-1,6-anhydro-6-deoxy- β -D-glucopyranose (14).—Obtained as described above from **7** (0.695 mmol); 0.192 g (84%); mp 124°C ; $[\alpha]_D -45.8^\circ$ (c 1.0, CHCl_3); R_f 0.31 (EtOAc); major rotamer (52%): ^1H NMR (CDCl_3): δ 5.74 (bs, 1 H, H-1), 4.93 (bs, 1 H, H-3), 4.88 (bs, 1 H, H-2), 4.72 (m, 1 H, H-5), 4.63 (bs, 1 H, H-4), 3.65 (m, 2 H, H-6a,6b), 2.20, 2.19, 2.17, 2.08 (4s, 12 H, CH_3CO); ^{13}C NMR (CDCl_3): δ 169.70, 169.15, 168.31, 167.82 (4 C, CH_3CO), 84.00 (C-1), 75.65 (C-5), 69.66 (C-4), 68.87 (C-3), 66.36 (C-2), 44.76 (C-6), 22.19, 20.78, 20.68, 20.56 (4 C, CH_3CO); minor rotamer (48%): ^1H NMR (CDCl_3): δ 5.51 (bs, 1 H, H-1), 4.91 (bs, 1 H, H-3), 4.69 (m, 1 H, H-5), 4.67 (bs, 1 H, H-4), 4.61 (bs, 1 H, H-2), 3.72 (dd, 1 H, $J_{5,6a}$ 6.6 Hz, $J_{6a,6b}$ 9.2 Hz, H-6a), 3.59 (d, 1 H, H-6b), 2.17, 2.16, 2.10, 2.08 (4s, 12 H, CH_3CO); ^{13}C NMR (CDCl_3): δ 169.94, 169.63, 168.76, 166.90 (4 C, CH_3CO), 83.88 (C-1),

74.88 (C-5), 69.26 (C-4), 68.78 (C-3), 68.31 (C-2), 46.32 (C-6), 22.09, 20.78, 20.68, 20.50 (4 C, CH₃CO). Anal. Calcd for C₁₄H₁₉NO₈ (329.30): C, 51.06; H, 5.82; N, 4.25. Found: C, 51.04; H, 5.94; N, 4.18.

6-Acetamido-2,3,4-tri-O-acetyl-1,6-anhydro-6-deoxy-β-D-galactopyranose (15).—Obtained as described above from **8** (0.695 mmol); 0.187 g (82%); mp 124 °C; [α]_D +22.1° (c 1.0, CHCl₃); *R*_f 0.30 (EtOAc); major rotamer (55%): ¹H NMR (CDCl₃): δ 5.66 (dd, 1 H, *J*_{1,2}, *J*_{1,3} 1.5 Hz, H-1), 5.33–5.30 (m, 1 H, H-3), 5.27 (dd, 1 H, *J*_{3,4} 5.4 Hz, *J*_{4,5} 4.3 Hz, H-4), 5.04 (dd, 1 H, *J*_{2,3} 1.9 Hz, H-2), 4.58–4.54 (m, 1 H, H-5), 3.77 (d, 1 H, *J*_{6a,6b} 8.8 Hz, H-6a), 3.61 (dd, 1 H, *J*_{5,6b} 5.9 Hz, H-6b), 2.18, 2.14, 2.09, 2.03 (4s, 12 H, CH₃CO); ¹³C NMR (CDCl₃): δ 169.83, 169.64, 169.46, 169.21 (4 C, CH₃CO), 84.09 (C-1), 73.72 (C-5), 70.94 (C-2), 67.34 (C-3), 64.70 (C-4), 44.31 (C-6), 22.57, 21.00, 20.76, 20.76 (4 C, CH₃CO); minor rotamer (45%): ¹H NMR (CDCl₃): δ 5.46 (dd, 1 H, *J*_{1,2}, *J*_{1,3} 1.7 Hz, H-1), 5.33–5.30 (m, 1 H, H-3), 5.27 (dd, 1 H, *J*_{3,4} 5.4 Hz, *J*_{4,5} 4.3 Hz, H-4), 4.77 (dd, 1 H, *J*_{2,3} 1.7 Hz, H-2), 4.54–4.52 (m, 1 H, H-5), 3.76 (d, 1 H, *J*_{6a,6b} 10.9 Hz, H-6a), 3.52 (dd, 1 H, *J*_{5,6b} 6.1 Hz, H-6b), 2.16, 2.11, 2.08, 2.04 (4s, 12 H, CH₃CO); ¹³C NMR (CDCl₃): δ 170.05, 169.21, 168.49, 167.15 (4 C, CH₃CO), 84.24 (C-1), 74.46 (C-5), 69.00 (C-2), 67.42 (C-3), 64.82 (C-4), 45.58 (C-6), 22.59, 20.99, 20.77, 20.77 (4 C, CH₃CO). Anal. Calcd for C₁₄H₁₉NO₈ (329.30): C, 51.06; H, 5.82; N, 4.25. Found: C, 51.20; H, 5.82; N, 3.98.

6-Acetamido-2,3,4-tri-O-acetyl-1,6-anhydro-6-deoxy-β-D-mannopyranose (16).—Obtained as described above from **9** (0.695 mmol); 0.189 g (82%); oily material; [α]_D –94.1° (c 1.0, CHCl₃); *R*_f 0.31 (EtOAc); major rotamer (57%): ¹H NMR (CDCl₃): δ 5.48 (dd, 1 H, *J*_{1,2} 2.5 Hz, *J*_{1,3} 1.4 Hz, H-1), 5.32 (ddd, 1 H, *J*_{2,3} 5.1 Hz, *J*_{3,4} 1.9 Hz, H-3), 5.24 (dd, 1 H, H-2), 4.84 (dd, 1 H, *J*_{4,5} 1.7 Hz, H-4), 4.68 (ddd, 1 H, *J*_{5,6a} 1.7 Hz, *J*_{5,6b} 6.8 Hz, H-5), 3.78–3.66 (m, 2 H, H-6a,6b), 2.18, 2.10, 2.08, 2.06 (4s, 12 H, CH₃CO); ¹³C NMR (CDCl₃): δ 169.84, 169.64, 168.41, 167.82 (4 C, CH₃CO), 84.62 (C-1), 75.45 (C-5), 72.06 (C-4), 67.85 (C-3), 67.75 (C-2), 45.64 (C-6), 22.78, 20.99, 20.69, 20.69 (4 C, CH₃CO); minor rotamer (43%): ¹H NMR (CDCl₃): δ 5.94 (dd, 1 H, *J*_{1,2}, 2.7 Hz, *J*_{1,3} 1.2 Hz, H-1), 5.35 (dd, 1 H, *J*_{2,3} 5.3 Hz, *J*_{3,4} 1.8 Hz, H-3), 5.16 (dd, 1 H, H-2), 4.83 (dd, 1 H, *J*_{4,5} 1.7 Hz, H-4), 4.71 (m, 1 H, H-5), 3.77–3.66 (m, 2 H, H-6a,6b), 2.18, 2.10, 2.08, 2.06 (4s, 12 H, CH₃CO); ¹³C NMR

(CDCl₃): δ 170.05, 169.25, 169.20, 167.05 (4 C, CH₃CO), 82.55 (C-1), 76.00 (C-5), 72.31 (C-4), 67.98 (C-3), 67.39 (C-2), 46.98 (C-6), 22.68, 21.30, 21.03, 20.95 (4 C, CH₃CO). Anal. Calcd for C₁₄H₁₉NO₈, H₂O (347.316): C, 48.41; H, 6.09; N, 4.03. Found: C, 48.12; H, 5.74, N, 3.91.

General procedure for the preparation of N-alkoxycarbonyl and N-acyl derivatives of the 2,3,4-tri-O-acetyl-6-amino-1,6-anhydro-6-deoxy-β-D-glucopyranose (17–20).—Alkyl chloroformate or acyl chloride (1.3 equiv.) was added at 0 °C to a soln of crude compound **10** [obtained from the salts **7** (0.695 mmol) as previously described] in C₅H₅N (5 mL). Stirring was maintained overnight; the soln was then concentrated and the residue was purified by column chromatography.

2,3,4-Tri-O-acetyl-1,6-anhydro-6-deoxy-6-methoxycarbonylamino-β-D-glucopyranose (17).—Obtained as described above from **10** and methyl chloroformate; the crude product was purified by column chromatography (2:1 EtOAc–petroleum ether); 0.212 g (88%); oily material; [α]_D –36.0° (c 1.0, CHCl₃); *R*_f 0.50 (2:1 EtOAc–petroleum ether); major rotamer (62%): ¹H NMR (CDCl₃): δ 5.48 (bs, 1 H, H-1), 4.87–4.65 (m, 4 H, H-2,3,4,5), 3.77 (s, 3 H, COOCH₃), 3.60 (m, 2 H, H-6a,6b), 2.17, 2.07, 2.07 (3s, 9 H, CH₃COO); ¹³C NMR (CDCl₃): δ 169.51, 169.20, 168.44 (3 C, CH₃COO), 153.12 (NCOO), 83.59 (C-1), 74.67 (C-5), 69.45 (C-4), 68.67 (C-3), 67.06 (C-2), 52.74 (COOCH₃), 45.25 (C-6), 20.67, 20.60, 20.37 (3 C, CH₃COO); minor rotamer (38%): ¹H NMR (CDCl₃): δ 5.57 (bs, 1 H, H-1), 4.87–4.65 (m, 4 H, H-2,3,4,5), 3.77 (s, 3 H, COOMe), 3.60 (m, 2 H, H-6a,6b), 2.17, 2.07, 2.07 (3s, 9 H, CH₃COO); ¹³C NMR (CDCl₃): δ 169.51, 169.20, 168.44 (3 C, CH₃CO), 153.12 (NCOO), 84.12 (C-1), 75.36 (C-5), 69.45 (C-4), 68.67 (C-3), 66.77 (C-2), 52.74 (COOCH₃), 45.25 (C-6), 20.67, 20.60, 20.37 (3 C, CH₃COO). Anal. Calcd for C₁₄H₁₉NO₉ (345.30): C, 48.69; H, 5.55; N, 4.06. Found: C, 48.36; H, 5.62, N, 3.94.

2,3,4-Tri-O-acetyl-6-allyloxycarbonylamino-1,6-anhydro-6-deoxy-β-D-glucopyranose (18).—Obtained as described above from **10** and allyl chloroformate; the crude product was purified by column chromatography (5:2 EtOAc–petroleum ether); 0.237 g (92%); oily material; [α]_D –38.6° (c 1.0, CHCl₃); *R*_f 0.70 (2:1 EtOAc–petroleum ether); major rotamer (63%): ¹H NMR (CDCl₃): δ 5.93 (m, 1 H, allyl CH=), 5.52 (bs, 1 H, H-1), 5.35 and 5.25 (m, 2 H, allyl CH₂=), 4.88–4.65 (m, 6 H, H-2,3,4,5, allyl CH₂), 3.60 (m, 2 H, H-6a,6b), 2.17,

2.07, 2.07 (3s, 9 H, CH_3CO); ^{13}C NMR (CDCl_3): δ 169.65, 169.34, 168.60 (3 C, CH_3COO), 152.39 (NCOO), 132.20 ($\text{CH}=\text{}$), 117.94 ($\text{CH}_2=\text{}$), 83.65 (C-1), 74.85 (C-5), 69.53 (C-4), 68.86 (C-3), 67.19 (C-2), 66.18 (CH_2O), 45.36 (C-6), 20.83, 20.74, 20.53 (3 C, CH_3COO); minor rotamer (37%): ^1H NMR (CDCl_3): δ 5.93 (m, 1 H, allyl $\text{CH}=\text{}$), 5.57 (bs, 1 H, H-1), 5.35 and 5.25 (m, 2 H, allyl $\text{CH}_2=\text{}$), 4.88–4.65 (m, 6 H, H-2,3,4,5, allyl CH_2), 3.60 (m, 2 H, H-6a,6b), 2.17, 2.07, 2.07 (3s, 9 H, CH_3COO); ^{13}C NMR (CDCl_3): δ 169.65, 169.34, 168.60 (3 C, CH_3COO), 152.39 (NCOO), 132.42 ($\text{CH}=\text{}$), 117.94 ($\text{CH}_2=\text{}$), 84.31 (C-1), 75.54 (C-5), 69.71 (C-4), 68.86 (C-3), 66.90 (C-2), 66.18 (CH_2O), 45.36 (C-6), 20.83, 20.74, 20.53 (3 C, CH_3COO). Anal. Calcd for $\text{C}_{16}\text{H}_{21}\text{NO}_9$ (371.335): C, 51.75; H, 5.70; N, 3.77. Found: C, 51.69; H, 5.79; N, 3.67.

2,3,4-Tri-O-acetyl-1,6-anhydro-6-deoxy-6-tetradecanoylamino- β -D-glucopyranose (19).—Obtained as described above from **10** and tetradecanoyl chloride; the crude product was purified by column chromatography (4:3 EtOAc–petroleum ether); 0.280 g (81%): mp 58–59 °C; $[\alpha]_{\text{D}} -26.0^\circ$ (c 1.0, CHCl_3); R_f 0.44 (1:1 EtOAc–petroleum ether); major rotamer (52%): ^1H NMR (CDCl_3): δ 5.76 (bs, 1 H, H-1), 4.94 (bs, 1 H, H-3), 4.88 (bs, 1 H, H-2), 4.73 (m, 1 H, H-5), 4.63 (bs, 1 H, H-4), 3.65 (m, 2 H, H-6a,6b), 2.34 (m, 2 H, CH_2CO), 2.20, 2.18, 2.07 (3s, 9 H, CH_3COO), 1.67 (m, 2 H, $\text{CH}_2\text{CH}_2\text{CO}$), 1.26 (m, 20 H, 10 CH_2), 0.88 (t, 3 H, J 6.4 Hz, CH_3CH_2); ^{13}C NMR (CDCl_3): δ 170.61, 170.15, 169.84, 168.28 (4 C, 3 CH_3CO , CH_2CO), 84.22 (C-1), 75.66 (C-5), 69.99 (C-4), 69.05 (C-3), 66.66 (C-2), 45.77 (C-6), 34.67, 31.81, 29.53–29.01, 25.19–24.29, 22.57 (12 CH_2), 20.81, 20.70, 20.63 (3 C, CH_3COO), 14.00 (CH_3CH_2); minor rotamer (48%): ^1H NMR (CDCl_3): δ 5.55 (bs, 1 H, H-1), 4.92 (bs, 1 H, H-3), 4.69 (m, 1 H, H-5), 4.68 (bs, 1 H, H-4), 4.61 (bs, 1 H, H-2), 3.70 (dd, 1 H, $J_{5,6a}$ 6.6 Hz, $J_{6a,6b}$ 9.2 Hz, H-6a), 3.58 (d, 1 H, H-6b), 2.34 (m, 2 H, CH_2CO), 2.21, 2.16, 2.07 (3s, 9 H, CH_3CO), 1.67 (m, 2 H, $\text{CH}_2\text{CH}_2\text{CO}$), 1.26 (m, 20 H, 10 CH_2), 0.88 (t, 3 H, J 6.4 Hz, CH_3CH_2); ^{13}C NMR (CDCl_3): δ 169.99, 169.69, 169.14, 168.75 (4 C, 3 CH_3CO , CH_2CO), 83.39 (C-1), 74.72 (C-5), 69.30 (C-4), 68.91 (C-3), 68.47 (C-2), 44.79 (C-6), 34.61, 31.81, 29.53–29.01, 25.19–24.29, 22.57 (12 CH_2), 20.81, 20.71, 20.52 (3 C, CH_3CO), 14.00 (CH_3CH_2). Anal. Calcd for $\text{C}_{26}\text{H}_{43}\text{NO}_8$ (497.61): C, 62.75; H, 8.71; N, 2.81. Found: C, 62.94; H, 8.87; N, 2.63.

2,3,4-Tri-O-acetyl-1,6-anhydro-6-deoxy-6-octanoylamino- β -D-glucopyranose (20).—Obtained as described above from **10** and octanoyl chloride; the crude product was purified by column chromatography (5:2 EtOAc–petroleum ether); 0.241 g (84%): mp 78–80 °C; $[\alpha]_{\text{D}} -28.3^\circ$ (c 1.0, CHCl_3); R_f 0.62 (5:2 EtOAc–petroleum ether); major rotamer (54%): ^1H NMR (CDCl_3): δ 5.76 (bs, 1 H, H-1), 4.93 (bs, 1 H, H-3), 4.88 (bs, 1 H, H-2), 4.72 (m, 1 H, H-5), 4.61 (bs, 1 H, H-4), 3.66 (m, 2 H, H-6a,6b), 2.33 (m, 2 H, CH_2CO), 2.19, 2.18, 2.06 (3s, 9 H, CH_3COO), 1.67 (m, 2 H, $\text{CH}_2\text{CH}_2\text{CO}$), 1.26 (m, 8 H, 4 CH_2), 0.88 (t, 3 H, J 6.4 Hz, CH_3CH_2); ^{13}C NMR (CDCl_3): δ 170.55, 170.06, 169.87, 168.36 (4 C, 3 CH_3CO , CH_2CO), 84.30 (C-1), 75.79 (C-5), 70.07 (C-4), 69.13 (C-3), 66.74 (C-2), 45.84 (C-6), 34.75, 31.65, 29.37, 29.05, 24.36, 22.59 (6 CH_2), 20.93, 20.83, 20.73 (3 C, CH_3CO), 14.05 (CH_3CH_2); minor rotamer (46%): ^1H NMR (CDCl_3): δ 5.55 (bs, 1 H, H-1), 4.90 (bs, 1 H, H-3), 4.72 (m, 1 H, H-5), 4.68 (bs, 1 H, H-4), 4.61 (bs, 1 H, H-2), 3.70 (dd, 1 H, $J_{5,6a}$ 6.6 Hz, $J_{6a,6b}$ 9.2 Hz, H-6a), 3.58 (d, 1 H, H-6b), 2.28 (m, 2 H, CH_2CO), 2.20, 2.16, 2.07 (3s, 9 H, CH_3CO), 1.67 (m, 2 H, $\text{CH}_2\text{CH}_2\text{CO}$), 1.26 (m, 8 H, 4 CH_2), 0.88 (t, 3 H, J 6.4 Hz, CH_3CH_2); ^{13}C NMR (CDCl_3): δ 170.06, 169.76, 169.20, 168.82 (4 C, 3 CH_3CO , CH_2CO), 83.50 (C-1), 74.83 (C-5), 69.42 (C-4), 69.04 (C-3), 68.60 (C-2), 44.87 (C-6), 34.69, 31.65, 29.22, 29.02, 25.27, 22.59 (6 CH_2), 20.93, 20.82, 20.64 (3 C, CH_3CO), 14.05 (CH_3CH_2). Anal. Calcd for $\text{C}_{20}\text{H}_{31}\text{NO}_8$ (413.455): C, 58.10; H, 7.56; N, 3.39. Found: C, 58.05; H, 7.72; N, 3.33.

1,6-Anhydro-6-deoxy-6-tetradecanoylamino- β -D-glucopyranose (21).—The product **19** (0.249 g, 0.50 mmol) was dissolved in dry MeOH (15 mL) and treated with a catalytic amount of NaOMe. After 16 h at room temperature, the mixture was concentrated and the residue was purified on a short column of silica-gel (6:1 EtOAc–EtOH); 0.154 g (83%): mp 97–98 °C; $[\alpha]_{\text{D}} -3.0^\circ$ (c 1.0, 1:1 CHCl_3 –EtOH); R_f 0.55 (6:1 EtOAc–EtOH); major rotamer (75%): ^1H NMR ($\text{C}_5\text{D}_5\text{N}$): δ 6.27 (bs, 1 H, H-1), 4.94 (bd, 1 H, $J_{5,6b}$ 6.5 Hz, H-5), 4.59 (m, 2 H, H-3,4), 4.18 (bs, 1 H, H-2), 4.05 (d, 1 H, $J_{6a,6b}$ 8.5 Hz, H-6a), 3.72 (dd, 1 H, H-6b), 2.35 (t, 2 H, CH_2CO), 1.77 (m, 2 H, $\text{CH}_2\text{CH}_2\text{CO}$), 1.19 (m, 20 H, 10 CH_2), 0.84 (t, 3 H, J 6.4 Hz, CH_3CH_2); minor rotamer (25%): ^1H NMR ($\text{C}_5\text{D}_5\text{N}$): δ 5.99 (bs, H-1), 4.92 (bd, 1 H, $J_{5,6b}$ 7.8 Hz, H-5), 4.59 (m, 2 H, H-3,4), 4.33 (bs, 1 H, H-2), 4.25 (d, 1 H, $J_{6a,6b}$ 10.4 Hz, H-6a), 3.90 (dd, 1 H, H-6b), 2.43 (t, 2 H,

CH_2CO), 1.77 (m, 2 H, $\text{CH}_2\text{CH}_2\text{CO}$), 1.19 (m, 20 H, 10 CH_2), 0.84 (t, 3 H, J 6.4 Hz, CH_3CH_2). Anal. Calcd for $\text{C}_{20}\text{H}_{37}\text{NO}_5$ (371.50): C, 64.66; H, 10.04; N, 3.77. Found: C, 64.81; H, 10.02; N, 3.72.

1,6-Anhydro-6-deoxy-6-octanoylamino- β -D-glucopyranose (22).—Prepared as described above from **21** (0.206 g, 0.50 mmol). The crude product was purified on a short column of silica-gel (4:1 EtOAc–EtOH); 0.121 g (84%): mp 69–70 °C; $[\alpha]_{\text{D}} -5.8^\circ$ (c 1.0, H_2O); R_f 0.35 (4:1 EtOAc–EtOH); major rotamer (75%): ^1H NMR ($\text{C}_5\text{D}_5\text{N}$): δ 6.31 (bs, 1 H, H-1), 4.99 (bd, 1 H, $J_{5,6b}$ 6.9 Hz, H-5), 4.62 (m, 2 H, H-3,4), 4.17 (bs, 1 H, H-2), 4.05 (d, 1 H, $J_{6a,6b}$ 8.4 Hz, H-6a), 3.71 (dd, 1 H, H-6b), 2.36 (t, 2 H, CH_2CO), 1.77 (m, 2 H, $\text{CH}_2\text{CH}_2\text{CO}$), 1.16 (m, 8 H, 4 CH_2), 0.79 (t, 3 H, J 6.4 Hz, CH_3CH_2); ^{13}C NMR ($\text{C}_5\text{D}_5\text{N}$): δ 170.74 (CH_2CO), 88.09 (C-1), 79.92 (C-5), 74.18, 72.71, 69.80 (3 C, C-2,3,4), 46.69 (C-6), 35.06, 32.08, 29.75, 29.59, 24.99, 23.00 (6 CH_2), 14.38 (CH_3CH_2); minor rotamer (25%): ^1H NMR ($\text{C}_5\text{D}_5\text{N}$): δ 6.00 (bs, 1 H, H-1), 4.96 (bd, 1 H, $J_{5,6b}$ 8.4 Hz, H-5), 4.59 (m, 2 H, H-3,4), 4.35 (bs, 1 H, H-2), 4.25 (d, 1 H, $J_{6a,6b}$ 10.3 Hz, H-6a), 3.89 (dd, 1 H, H-6b), 2.45 (t, 2 H, CH_2CO), 1.77 (m, 2 H, $\text{CH}_2\text{CH}_2\text{CO}$), 1.19 (m, 8 H, 4 CH_2), 0.84 (t, 3 H, J 6.4 Hz, CH_3CH_2); ^{13}C NMR ($\text{C}_5\text{D}_5\text{N}$): δ 169.42 (CH_2CO), 88.35 (C-1), 79.07 (C-5), 74.18, 72.87, 72.22 (3 C, C-2,3,4), 46.46 (C-6), 35.06, 32.08, 29.75, 29.59, 25.65, 23.00 (6 CH_2), 14.38 (CH_3CH_2). Anal. Calcd for $\text{C}_{14}\text{H}_{25}\text{NO}_5$ (287.35): C, 58.51; H, 8.77; N, 4.87. Found: C, 58.74; H, 8.82; N, 4.83.

References

- [1] H. Paulsen and K. Todt, *Angew. Chem.*, 77 (1965) 589.
- [2] H. Paulsen and K. Todt, *Chem. Ber.*, 99 (1966) 3450–3460; H. Paulsen and K. Todt, *Chem. Ber.*, 100 (1967) 512–520.
- [3] W. Meyer zu Reckendorf and N. Wassiliadou-Micheli, *Chem. Ber.*, 101 (1968) 2294–2301.
- [4] H. Paulsen and K. Todt, *Adv. Carbohydr. Chem.*, 23 (1968) 115–232.
- [5] M.A. Pradera, D. Olano, and J. Fuentes, *Tetrahedron Lett.*, 36 (1995) 8653–8656.
- [6] J. Fuentes, D. Olano, and M.A. Pradera, *Tetrahedron: Asymmetry*, 8 (1997) 3443–3456.
- [7] D. Lafont, P. Guilloux, and G. Descotes, *Carbohydr. Res.*, 193 (1989) 61–73.
- [8] D. Lafont, P. Boullanger, F. Carvalho, and Ph. Vottero, *Carbohydr. Res.*, 297 (1997) 117–126.
- [9] H. Staudinger and J. Meyer, *Helv. Chim. Acta*, 2 (1919) 635–646.
- [10] A. Messmer, I. Pintér, and F. Szegő, *Angew. Chem.*, 76 (1964) 227–228.
- [11] I. Pintér, J. Kovács, A. Messmer, A. Kálmán, G. Tóth, and B.K. Lindberg, *Carbohydr. Res.*, 72 (1979) 289–296.
- [12] J. Kovács, I. Pintér, F. Szegő, G. Tóth, and A. Messmer, *Acta Chim. Acad. Sci. Hung.*, 101 (1979) 7–16.
- [13] V. Zsoldos-Mády, I. Pintér, P. Sándor, and A. Messmer, *Carbohydr. Res.*, 281 (1996) 321–326.
- [14] Z. Györgydeák, L. Szilágyi, and H. Paulsen, *J. Carbohydr. Chem.*, 12 (1993) 139–163.
- [15] V. Maunier, P. Boullanger, and D. Lafont, *J. Carbohydr. Chem.*, 16 (1997) 231–235 and ref. therein.
- [16] D. Lafont, P. Boullanger, O. Cadas, and G. Descotes, *Synthesis*, (1989) 191–194 and refs. therein.
- [17] H. Driguez, J.C. McAuliffe, R.V. Stick, D.M.G. Tilbrook, and S.J. Williams, *Aust. J. Chem.*, 49 (1996) 343–348 and refs. therein.
- [18] E. Hardegger and R.M. Montavon, *Helv. Chim. Acta*, 29 (1946) 1199–1203.
- [19] H. Ohle and H. Thiel, *Chem. Ber.*, 66 (1933) 525–532.
- [20] F.D. Tropper, F.O. Anderson, S. Braun, and R. Roy, *Synthesis*, (1992) 618–620.
- [21] Z. Györgydeák and H. Paulsen, *Liebigs Ann. Chem.*, (1977) 1987–1991.
- [22] R.G.S. Ritchie, N. Cyr, and A.S. Perlin, *Can. J. Chem.*, 54 (1976) 2301–2309.
- [23] T.B. Grindley and R. Thangarasa, *Carbohydr. Res.*, 172 (1988) 311–318.
- [24] J. Kovács, I. Pintér, and A. Messmer, *Carbohydr. Res.*, 141 (1985) 57–65.
- [25] J. Kovács, I. Pintér, A. Messmer, G. Tóth, and H. Duddeck, *Carbohydr. Res.*, 166 (1987) 101–111.
- [26] A.A. Bothner-By, R.L. Stephens, and J. Lee, *J. Am. Chem. Soc.*, 106 (1984) 811–813.
- [27] D. Lafont and P. Boullanger, *J. Carbohydr. Chem.*, 11 (1992) 567–586.
- [28] W.A. Szarek, S. Wolfe, and J.K.N. Jones, *Tetrahedron Lett.*, 38 (1964) 2743–2750.
- [29] M.L. Wolfrom, J.L. Minor, and W.A. Szarek, *Carbohydr. Res.*, 1 (1965) 156–165.
- [30] H. Paulsen and K. Todt, *Chem. Ber.*, 100 (1967) 3385–3396, 3397–3404.
- [31] P. Lecomte du Nouÿ, *J. Gen. Physiol.*, 1 (1918) 521–524.
- [32] W.D. Harkins and H.F. Jordan, *J. Am. Chem. Soc.*, 52 (1930) 1751–1772.
- [33] Th. Böcker and J. Thiem, *Tenside Surf. Det.*, 26 (1989) 318–324.
- [34] A.R. van Buuren and H.J.C. Berendsen, *Langmuir*, 10 (1994) 1703–1713.
- [35] Z. Györgydeák and L. Szilágyi, *Liebigs Ann. Chem.*, (1987) 235–242.